PRELIMINARY STUDIES ON THE ESTIMATION

OF LARVAL POPULATIONS

OF THE LARCH SAWFLY, PRISTIPHORA ERICHSONII (HTG.)

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ABSTRACT

The seasonal development of eggs and larvae of the larch sawfly are shown for 1957 and 1958. The general trends were similar for both years although the position of a minor peak differed between the two years.

A comparison of cumulative adjusted oviposition curves with numbers of egg scars suggests that rate of development in the insectary was comparable to that under field conditions.

The percentages of mortality estimated from cumulative adjusted population figures were: egg to first instar - 27.0 per cent; first to second instar - 1.3 per cent; second to fourth instar - 32.9 per cent; fourth to fifth instar - 37.0 per cent. These estimates compared favourably to figures derived from alternative methods of estimating mortality.

The magnitude and nature of the variation in insect numbers is discussed briefly.

1. INTRODUCTION

Sampling techniques have been developed for estimating larch sawfly populations at various stages in the insect's life cycle, but not for estimating feeding larval populations. Most of the mortality during the insect's life cycle occurs in the interval between the oviposition of eggs and the completion of larval feeding (Greham, 1956; Ives, 195h; Turnock, 1956). An attempt is therefore being made to develop a sampling method for estimating mortality during this period.

The most obvious approach to the problem of evaluating mortality is to tag a series of shoots containing eggs of the larch sawfly, then make periodic examinations and record the numbers and stages of larvae present. In practice, however, this is impractical. It is virtually impossible to obtain accurate counts of the early-instar larvae on the foliage, since they are gregarious feeders. Furthermore, colonies of larvae tend to split up and wander, especially during the fifth instar, so that a decrease in numbers may be due to wandering of some individuals. Confinement of the larvae by tanglefoot bands or other means might prevent insect predators from reaching them. The first of these difficulties could be circumvented by tagging a large number of shoots and estimating the number of insects present by sampling a number of the tagged shoots at each examination. However, this would not overcome the problem of larval wandering.

Assessment of mortality by observing tagged colonies was abandoned in favor of periodic sampling of whole branches. A number of difficulties are encountered in this approach also, as will be seen in this preliminary report covering work done in 1957 and 1958. The abundance of the various stages in the field at any given point in time is relatively easy to determine, since no particular mechanical difficulties were encountered. However, such figures do not give a true picture of the total number in each stage throughout the season. Eggs and fifth-instar larvae remain in their respective stages for a considerable period of time. Samples taken every four days will, in effect, sample a portion of these populations at least twice. The other four larval instars may or may not last less than four days, depending upon temperature, so that some of these may not have been sampled at all. For example, larvae in the first instar at one sampling period may be in the third instar when the next sample is taken four days later. These larvae will have formed part of the second-instar population for the period under consideration but will not be included in estimates based on field collections only. For this reason it was considered necessary to conduct a rearing program in conjunction with the sampling program, to provide estimates of the number of insects missed or sampled twice. In the present study it has been assumed that the rate of development under insectary conditions is the same as under field conditions, although the validity of this assumption will have to be tested.

Several approaches are available for estimating mortality. These are: (1) cumulative population totals; (2) hatch records; (3) association of larval colonies with shoots containing oviposition scars; and (4) comparison of field collections with insectary rearings of previous collections. None of these methods is completely satisfactory, as will be noted in the following discussion.

2. METHODS

The basic method used in randomizing the selection of branches was the same as in egg sampling (Ives, 1955). The 1957 sample consisted of two whole branches selected at random from below 30 feet on each of five trees. The branches were removed with pole pruners fitted with a clamp held by a locking device permitting the branches to be lowered gently (Fig. 1). Samples were taken every four days from a different random group of five trees. Sampling commenced June 10 and ended August 9. Only large branches were sampled and completely defoliated branches were rejected. A large net was periodically attached to the pole pruners to determine if larvae were being lost. Total shoots, scarred shoots, foliated length

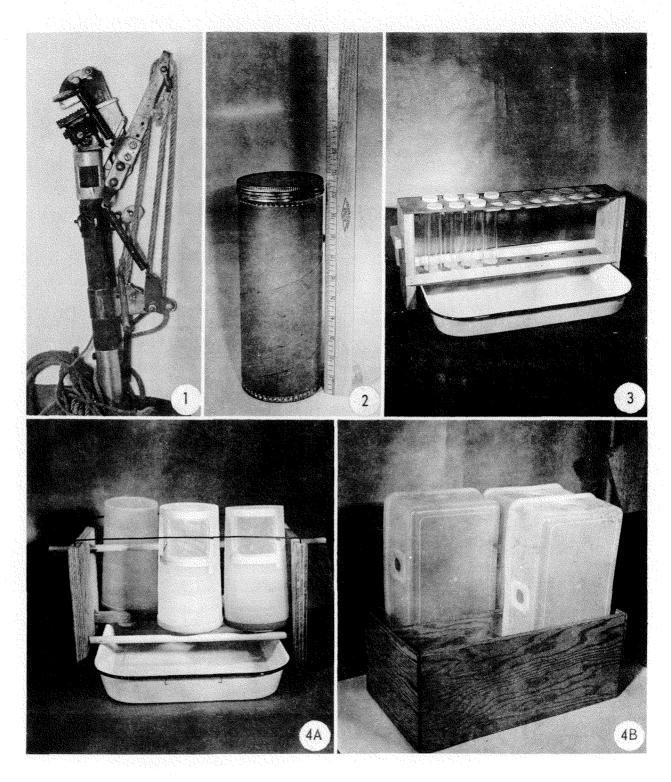
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and foliated width were recorded for each branch. Separate containers were used for packaging shoots containing eggs, shoots with egg scars not associated with larvae and larvae not associated with shoots. Each colony of larvae that was associated with a shoot was placed in a container with the respective shoot.

Upon return to the laboratory the shoots containing eggs were placed in water in small vials which in turn were placed in 4-inch shell vials stoppered with a cotton plug. The shoots were examined in four days and the number of larvae for each shoot recorded. Eggs that had not hatched in this period of time were reared until hatching occurred or until mortality could be assumed. After hatching was complete, the scars in each shoot were counted and recorded. All larvae were killed and counted by instar upon return to the laboratory, and records kept of the numbers of scars in associated shoots. Larvae or scarred shoots that could not be associated with one another were recorded separately, as were shoots containing scars but showing little or no evidence of larval feeding.

Several aspects of this procedure were modified in 1958. The sample size was increased to 20 branches, one from each of 20 trees. To facilitate sampling, the trees were sampled serially throughout the plot. Samples were again taken every four days, commencing June 11 and ending August 22. After the first sample, half of the sample was overlapped with the previous sample, i.e. the first sample consisted of branches from trees 1 to 20, the second of branches from trees 11 to 30, and so on. The branches in the overlapping half were sampled from the same height as the branches previously sampled but from opposite sides of the crown. The branches in the other half of the sample were selected at random from the crowns of **the trees** and determined the position of the branches in the overlapping half of the succeeding sample. Completely defoliated branches were retained in the sample. Field data were recorded as before, and field techniques were the same except that large screw top cardboard cans (Fig. 2) were used for transferring larvae from the field to the insectary.

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Figs. 1 - 4 1. Pole pruner head modified for larval sampling. 2. Screw top cardboard container for transporting larvae from the field to the insectary. 3. Rearing cages used for determining percentage hatch. 4. Plastic larval rearing cages: (a) small cages used for rearing early instar larvae: and (b) large cages used for rearing large colonies and fifth instar larvae. (Cut end of branches in plastic cages immersed in water-filled jars suspended from wooden box). Accompanying laboratory techniques were modified considerably. Special rearing containers (Fig. 3) were made for determining percentage of hatch, because conditions in the shell vials created high humidity which was unfavorable for larval development and the dead unfed larvae were difficult to count. All larvae were reared in the insectary in plastic containers (Fig. 4) until the next sampling period. Mixed groups were counted as accurately as possible before rearing, but homogeneous groups were counted only when the larvae were killed four days later. This procedure least disturbed the larvae and also gave maximum accuracy. Fifthinstar larvae were reared in cages containing a small amount of moist sphagnum moss. This provided a method for determining if larvae had finished feeding, since any found crawling in the moss were assumed to be ready to spin cocoons.

Adjusted population estimates, based on field collections and insectary rearings, were calculated for each sample from the formula

$$P_{k_{i}} = \sum_{j=1}^{n} X_{k_{ij}} - \sum_{j=1}^{n} Y_{k_{ij}} + \sum_{j=1}^{n} Z_{k_{ij}}$$

subject to the condition that $P_{k_i} \ge 0$ where P_{k_i} is the adjusted population th for the k stage of the insect (K=1, 2, -, 6) at the i sampling period; n is the number of branches in each sample; $X_{k_{i_j}}$ is the observed number of insects in the k stage on the j branch of the i sample; $Y_{k_{i_j}}$ is the number of insects from the j branch of the (i-1) sample remaining in the k stage during the period between collection of the $(i-1)^{t_1}$ and i samples; and $Z_{k_{i_j}}$ is the number of insects from the j branch of the (i-1) sample passing from the (k-1) to the (k+1) stage during the period between collection of the (i-1) samples.

The formula therefore estimates the number of insects that have entered or passed

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through any given stage in the 4-day period prior to the collection of any sample. It makes no provision for differential mortality between laboratory and field conditions.during the period under consideration. The conditions of non-negativity was imposed to make allowance for sampling variation or differential mortality that could result in negative answers.

3. RESULTS

The work in 1957 was largely confined to developing techniques, mainly removal of branches from the trees without losing too many larvae. Rearing techniques were not satisfactory and with the exception of egg hatch records will not be discussed.

The results to date are considered indicative only, and the current presentation will be largely confined to graphs and tabulations, with a minimum of statistical analysis.

3.1 Seasonal Development

The observed seasonal distribution of eggs and larvae for 1957 and 1958 and the adjusted distributions where estimates of these are available are shown in Appendix Tables 1 and 2. Negative answers are shown in parenthesis. The **same** distributions are shown in Figs. 5 and 6.

The frequency histograms for number of eggs provide a good illustration of how misleading field collections can be in indicating peak increments to insect populations. The field collections indicate peak oviposition for the four-day period prior to June 30, 1957 and July 9, 1958, a difference of nine days. Adjustment to allow for unhatched eggs shows that the peaks were probably June 26 and July 1, respectively. Nearly half of the time difference in apparent oviposition peaks was therefore due to slower development of the eggs in 1958.

The 1958 histograms for second-, third- and fourth-instar larvae illustrate

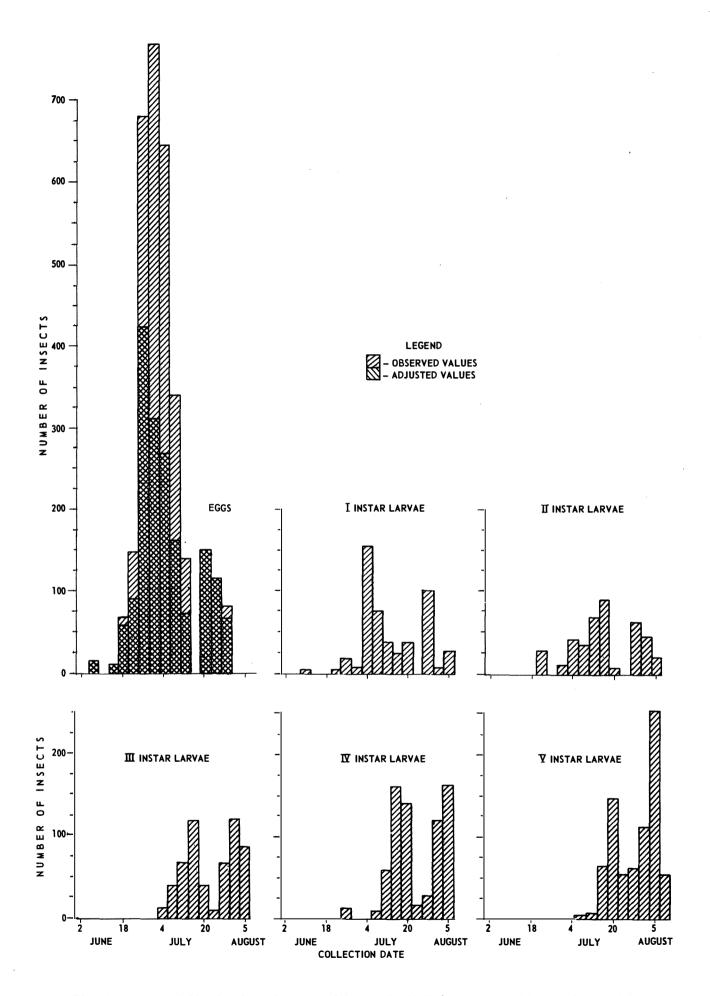


Fig. 5. Seasonal distribution of eggs and larvae in 1957 (number per 10-branch sample).

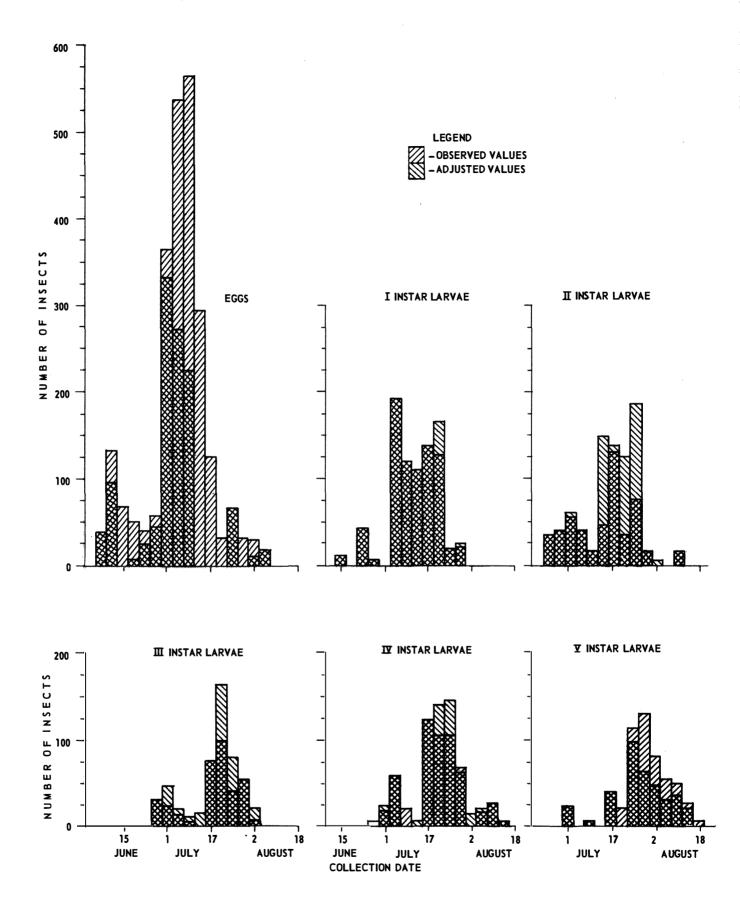


Fig. 6. Seasonal distribution of eggs and larvae in 1958 (number per 20-branch sample).

that a considerable number of larvae in these instars are missed entirely even when sampling is conducted every four days. The histograms for first-instar larvae should probably show a higher proportion of adjustment, but a flaw in technique may have prevented this. The histogram for fifth-instar larvae demonstrates that sampling every four days in effect samples a portion of the larvae in this stage twice.

Except for the slight differences in peak oviposition periods, the general trends for development are similar for 1957 and 1958. In 1957 there was a main peak in oviposition, followed by a secondary peak about three weeks later. This bimodality persisted throughout the larval instars, but because no adjusted population estimates are available it is difficult to determine the relative magnitudes of these peaks. In 1958 the main oviposition period occurred at about the same time as in 1957, but was preceded by a minor peak. The latter persisted through all larval instars. Although the samples are very variable at the present stage of development it appears that they are capable of giving a reasonably accurate picture of development periods for the different immature stages.

3.2 Cumulative oviposition curves

The total oviposition occurring prior to any point in time can be estimated by counting the total egg scars on the branches sampled. Comparison of these values with the cumulative totals of the adjusted oviposition estimates

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During periods where the work load was heavy on sampling days some of the hatch counts were made after less than four days of rearing. Some of these larvae would undoubtedly have entered the second instar had the rearing been continued for the full four days. This defect will be corrected in future work.

provides a check for gross errors in the latter. The results of this comparison are shown in Figs. 7 and 8 for 1957 and 1958 respectively. The same comparisons are shown in Appendix Table 3. The high variation between samples in the number of eggs makes an accurate check impossible, but the cumulative totals of adjusted values appear slightly higher than they should. Examination of egg clusters from the previous sample was delayed until after the next sample had been completed. When larvae and eggs were abundant this resulted in an extension of several hours beyond the four day period. This extension would probably be sufficient, especially on a hot day, to allow enough additional hatching to account for the discrepancies noted. Examination of egg clusters while field samples are being taken should eliminate this bias. Since the difference noted was small the foregoing explanation is probably correct, but similar effects would occur if the rate of development was more rapid in the insectary than in the field.

3.3 Estimation of mortality

3.3.1 Cumulative population totals

The adjusted values for eggs and larvae, as shown in Fig. 6, were simply added to give the cumulative totals shown in Fig. 9.

An inconsistency due to sampling variation is apparent in the population fourthestimates because there were more/than third-instar larvae. It has been assumed that there was no mortality between the third and fourth instars, and the pooled values have been used as the estimate of the number in the fourth instar. On this basis the apparent mortality, in percentages, based on the cumulative number of insects per 20 branches was as follows: egg to first instar - 27.0; first to second instar - 1.3; second to fourth instar - 32.9; fourth to fifth instar - 37.0. The corresponding percentages for real mortality were: 27.0, 1.0, 23.7 and 17.9. It therefore appears that the main periods where mortality occurred were:

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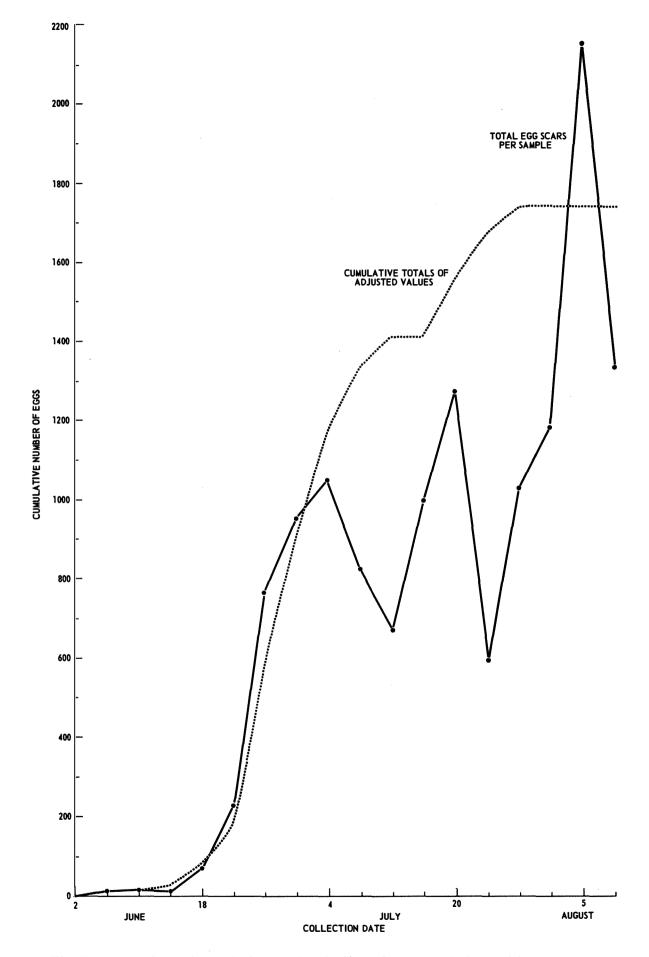


Fig. 7. Comparison of cumulative totals of adjusted egg populations with numbers of egg scars per sample (1957 data).

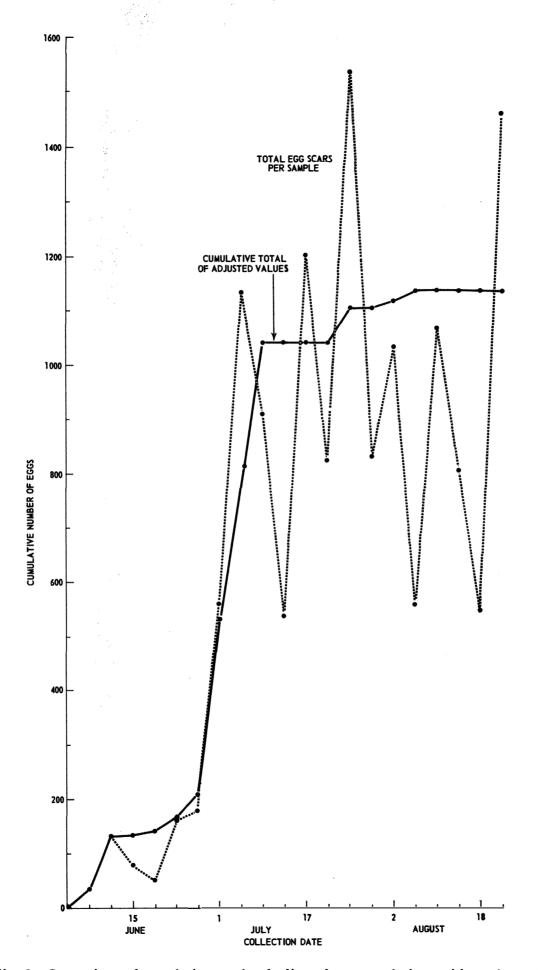


Fig. 8. Comparison of cumulative totals of adjusted egg populations with numbers of egg scars per sample (1958 data).

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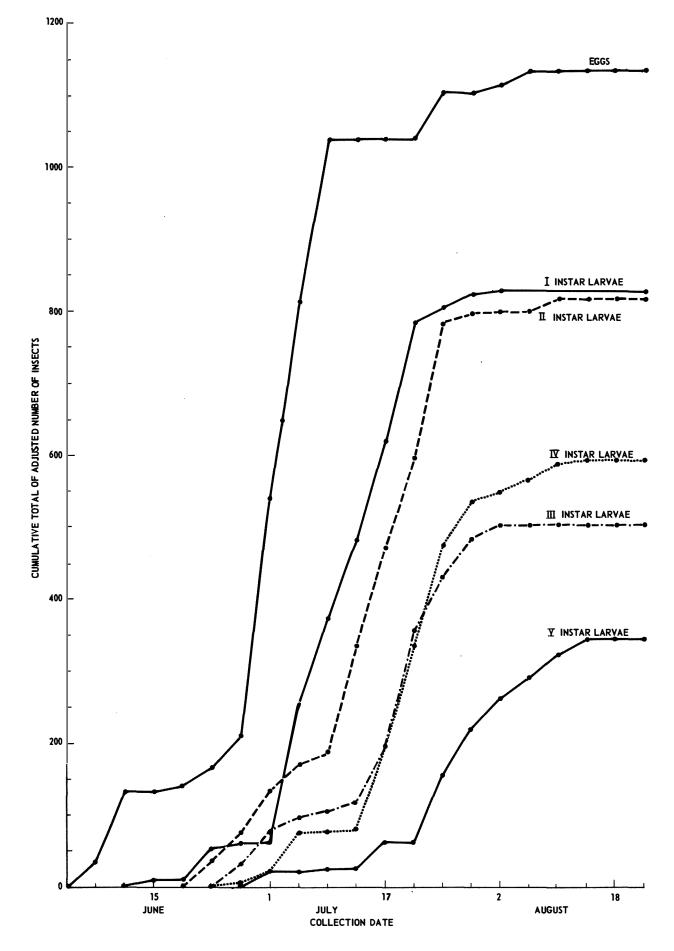


Fig. 9. Cumulative totals of adjusted population estimates for eggs and larvae (1958 data).

(1) between the egg stage and the middle of the first instar; (2) between the middle of the second and middle of the third instars; and (3) between the middle of the fourth and middle of the fifth instars. It should be emphasized, however, that these conclusions are based on data which is very variable and may therefore be erroneous.

3.3.2 Hatch records

Shoots containing eggs were reared to obtain records of hatching. To determine if insect predators were attacking the eggs, the records were divided into two groups based on the time interval between collection and hatching, those that had hatched or were hatching in four days and those that took longer (Appendix Tables 4 and 5). The percentages of hatching in the two groups were 78.9 and 93.4 in 1957 and 77.7 and 83.2 in 1958. These differences probably can be attributed to insect predation, as rearing conditions were identical. The differences do not represent the total insect predation, for some of the first group would normally have been exposed to predation for up to four days longer, and some of the second group had also been exposed to predation for a number of days before collection. Furthermore, if all of the eggs in a shoot had been destroyed it would not be included, as it was impossible to determine when the eggs had been laid. The differences probably represent approximately half of the total predation since the 4-day period is about half of the incubation period.

Early in the season it was usually possible to determine if there had been any feeding on or near a scarred shoot. If there was little or none the eggs or newly hatched larvae may have been destroyed by insect predators. A complete tally of all oviposition scars was therefore made, covering the period when it was possible to associate most of the scarred shoots with colonies of larvae (Appendix Table 6). Assuming that predators had destroyed all the eggs in the column headed

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"apparently predatorized", the percentages of losses are not large, amounting to 8.9 and 4.5 for 1957 and 1958, respectively. However, if they are added to the differences between the hatch figures for the two groups the amount of mortality that may be due to insect predators is 23.4 per cent in 1957 and 10.0 per cent in 1958. The percentage of mortality in the "not hatched in four days" group was 16.8 in 1958. Some of this mortality is probably due to insect predators, the remainder to infertility, failure of larvae to escape from egg slits and other causes. Adding the 1958 figures of 10.0 and 16.8 gives a figure (26.8) that is in very close agreement with the mortality figure (27.0) for the interval between the egg and first instar larva, as calculated from the cumulative number of insects.

3.3.3. Associated larvae and egg scars

The estimates of mortality based on association between larvae and egg scars have one major weakness. If all of the larvae in a colony are destroyed no estimate is available. The chances of this happening should increase with time and one would expect a greater under-estimation for later instars than for earlier instars. Another weakness is that no association is usually possible for fifthinstar larvae because of larval wandering. Nonetheless, the method does serve as a check on other mortality estimates by providing a lower bound. The number of scars and numbers of associated larvae for the first four instars for 1957 and 1958 are shown in Appendix Tables 7 and 8. The percentages "pseudo-real" mortality for 1958 are 38.6, 30.0, 43.9 and 50.3 for instars I, II, III and IV respectively. (The mortality has been called "pseudo-real" because it approaches the definition of real mortality but errs by an unknown amount because the eggs from colonies in which all larvae have been destroyed are not included.) There is another anomaly here which also occurred in 1957: the mortality to the second instar is less than that to the first instar. This anomaly may be due to sampling variation. If so,

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the best estimate of the mortality to the first instar is the pooled data for the first and second instars. Comparison of this pooled result for 1958 (35.8) with the cumulative insect estimate (27.0) indicates a disagreement of 8.8 per cent. However, if there is a mechanical source of error² in the estimates of mortality to the first instar, and in fact the true mortality to the second instar is in the neighbourhood of 30 per cent, the appropriate comparison is 27.0 plus 1.0 or 28.0 per cent and the two estimates are in reasonably close agreement.

Assuming 30.0 per cent to represent the mortality to the second instar, association between shoots and larvae gives an estimate of 13.9 per cent mortality between the second and third instars and a further 6.4 per cent between the third and fourth instars in 1958. The total "pseudo-real" mortality to the fourth instar, 50.3 per cent, is in close agreement with the real mortality estimated from the cumulative adjusted population figures, 51.6 per cent. It therefore appears that fewer colonies were completely destroyed than had been anticipated, although this agreement could be due to fortuitous circumstances.

3.3.4 Collections versus insectary rearings

Estimation of mortality by comparing the number of insects in each instar reared from the previous collection, with the number collected from the field has a number of disadvantages. The mortality is not estimated for any fixed number of instars, but for the interval between the collection of the two samples. The estimate is also subject to bias if the rate of insectary development differs from that in the field. Sampling variation also may greatly affect the estimates, as shown in Appendix Table 9, where fewer second- and fourth-instar larvae occurred in

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Such an error is quite possible. Larvae were not counted (in 1958) until the conclusion of rearing. If dead first-instar larvae were mistaken for cast skins, or missed, the results would over-estimate mortality to the first instar.

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the reared population than in the field population.

However, no check on the mortality between the fourth and fifth instars is available from other sources. Both this estimate and the one to be checked are based on insectary rearings, but the approach is different. The apparent mortality for the 4-day period prior to collection for the fifth-instar larvae is estimated as 36 per cent. This period approximately covers the late fourth and early fifth instars. The estimate therefore compares favorably with that of 37 per cent apparent mortality between the mid-fourth and mid-fifth instars as estimated from the cumulative adjusted population figures.

3.4 Sampling variation

Sampling variation has not been discussed in the previous section because the sample sizes were small and confidence intervals would be too wide to be of any value. However, examination of the variability will prove helpful in planning future work.

The 1957 data provides some information on the relative magnitudes of inter- and intra-tree variability, although inclusion of small and defoliated branches would undoubtedly have increased these variations (Appendix Table 10). The intra-tree variation, with some exceptions, is as large as the inter-tree variation for all stages sampled. Stratification of the sample by crown level and crown class has been shown to reduce the intra-tree variation in number of scarred shoots (Ives, 1955). Such stratification should also prove useful in larval sampling, but all crown levels probably could not be sampled from each of the trees since the number of trees would then be too small. Each stratum could be constructed to include two branches from one crown level of a group of trees having a similar crown class. In this way all crown levels and crown classes could be represented in each sample, although no estimate of inter-crown level or inter-crown class variability

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would be available. However, the error mean square should be reduced appreciably.

The 1958 data provides information on the variances and covariances for samples taken from the same trees at 4-day intervals (Appendix Table 11). With a few exceptions, the larval stages are apparently too transient to be able to demonstrate any appreciable continuity between successive samples, even when these samples consist of paired branches. The eggs and total scars show that some continuity is achieved by overlapping the samples, but the results are inconsistent. Mechanical difficulties would likely be encountered if two branches from a crown level were overlapped with a preceding sample of two branches (i.e. it would be difficult to ensure that removing the first two branches would not injure or dislodge larvae on branches to be included in the second sample). It therefore appears that the gain in continuity does not justify overlapping the samples. Independent samples would also simplify the estimation of variances, since no covariances would be involved between successive samples.

A further aspect of variation that is of concern in estimating larval populations is the amount of variation at different population levels. The available data are rather inadequate to determine the relationship between variability and population density, since only two years' collections are available and the populations were not high. However, examination of the data shows clearly that the variation increases as the population increases for all stages sampled in this project (Fig. 10).

The relationship between mean and standard deviation appears to be linear, although the egg data for 1958 suggests the possibility of a curvilinear relation. The accuracy of estimates at different population levels is roughly proportionate to the mean for the range in densities encountered. If this relationship holds over a wider range it will simplify the estimation of larval populations, since a fixed sample size can be used with equal efficiency throughout the season.

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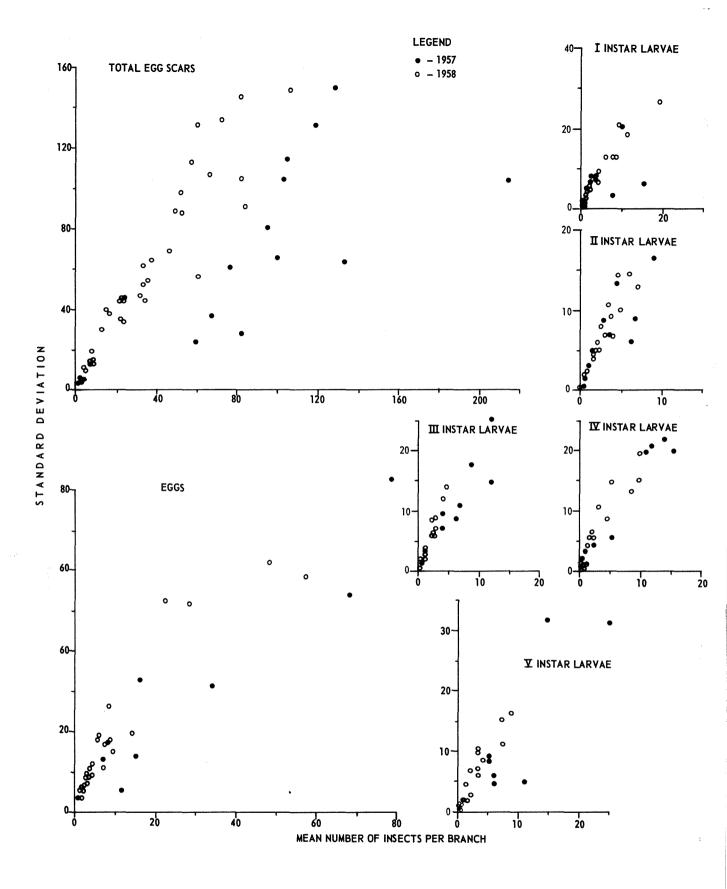


Fig. 10. Relationship between means and standard deviations in 1957 and 1958.

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Ives, W. G.	н.	1954.	Population studies of the larch sawfly <u>Pristiphora</u> erichsonii (Hartig). Tech. Report 1954. Forest Biology Laboratory, Winnipeg, Man.
Ives, W. G.	H.	1955.	Estimation of egg populations of the larch sawfly, Pristiphora erichsonii (Hartig). Can. Jour. Zool. 33: 370-388.
Turnock, W.	J	1956.	Preliminary life tables for the larch sawfly. Interim Tech. Report 1955 -7. Forest Biology Laboratory, Winnibeg, Man.

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5. APPENDIX

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Seasonal Distribution of Eggs and Larvae in 1957 (number per 10-branch sample)

ate	Eg Observed	Adjusted	I Instar	II Instar	III Instar	IV Instar	V Instar
une 2*	0	0	0	0	0	0	0
6	15	15	0	0	0	0	0
10	0	0	2	0	0	0	0
1/4	11	11	0	0	0	0	0
18	68	57	0	0	0	0	0
22	158	90	3	28	0	0	0
26	681	423	16	0	0	11	0
30	783	311	6	10	0	0	0
uly 4	646	267	154	41	12	0	Ō
8	339	161	74		39	6	Ţ
12	139	73	36	34 68	67	55	Ĵ,
16	0	Ó	22	89	118	158	62
20	149	149	35	6	40	138	146
24	114	114	0	0	8	12	
28	81	66	100	62	64	24	52 61
ug. 1	0	0	6	45	121	117	110
ς Γ	õ	õ	26	16	86	159	252
0	Ő	õ	0	0	0	1	52

Date		Egg Observed		I L Observed	nstar Adjusted		nsta r Adju ste d		Instar Adjusted		nsta r Adjusted	V I Observed	nstar Adjusted
June	3*	0	0	0	0	0	0	0	0	0	0	0	0
	7*	37	37	0	0	0	0	0	0	0	0	0	Ō
]	11	133 67	96	0	0	0	0	0	0	0	0	0	Ō
]	15	67	0(-40)	11	11	0	0	0	0	0	0	0	Ō
]	19	51	7	0	0	0	0	0	0	0	0	0	0
2	23	41	25	42	42	34	34	0	0	0	0	0	0
	27	57	144	6	6	39	39	30	30	3	3	0	0
July	j	365	332	0	0	55	59	23	45	14	19	21	21
	5	537	273	193	193	41	40	13	20	54 18	53	0	0(-21)
_	9	565	226	120	120	15	171	5	10	18	0(7)	3	<u>`</u> 3 ´
	13	294	0	111	\mathbf{m}	45	147	0	13	0	3	0	0(-3)
	17	126	0(-63)	138	138	130	138	75	75	119	119	38	38
2	21	30 66	0(-39)	127	165	36	126	9 8	162	103	137	18	0(-12)
	25		66	18	18	75	185	39 53	77	103	JJ13	109	` 94 ´
	29	29 28	0(-7)	20	22	16	16	53	53	64	61	127	63
Aug.	2		n	0	3	0	4	6	20	1	13	78	44
-	0	18	18	0	0	0	0	0	0	15	17	52	29
	LO	0	0	0	0	16	16	0	0	23	23	47	33
	4	0	0	0	0	0	0	0	0	3	2	24	21
Ţ	.8 22	0	0	0	0	0	0	0	0	0	0	4	0(-9)
2	22	0	0	0	0	0	0	0	0	0	0	Ó	0(-1)

Seasonal Distribution of Eggs and Larvae in 1958 (number per 20-branch sample). Negative answers for adjusted values are shown in parenthesis.

*Interpolated from June 11 sample.

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Comparison of Cumulative Totals of Adjusted Egg Populations with Numbers of Egg Scars per Sample

	1957			1958	
	Cumulative Adjusted	Number of egg		Cumulative Adjusted	Number of egg
Date	egg population	scars per sample	Date	egg population	scars per sample
June 2	2 0	0	June 3	0	0
	5 15	15	7	37	. 37
10	0 15	15	11	133	133
	4 26 8 83	11	15	133	79
גר 12	8 83	68	19	140	51
22	2 173	227	23	165	162
26	5 596	763	27	209	182
30		952	July 1	541	560
July 1	1174	1052	5	814	1132
3		823	9	1040	908
12		671	13	1040	537
10		99 7	17	1040	1203
20		1276	21	1040	823
21	1671	592	25	1106	1538
28		1027	29	1106	830
		1180	Aug. 2	1117	· 1033
ļ	L 1737 5 1737 9 1737	2140	<u> </u>	1135	558
9	9 1737	1330	10	1135	1064
			14	1135	807
•			18	1135	546
			22	1135	1459

1957 Hatch Records

Eggs na	ggs hatched (or natching) in 4 days						bggs not nationed in 4 days						
Date col- lected	No. egg scars	No. larvae	Date col- lected	No. egg scars	No. larvze		Date col- lected	No. egg scars	No. larvae	Date co l- lected	No. egg scars	No. larvae	
26/VI 30/VI	27 27 38 18 7 14 33 22 19 24 26 28 21 30 15 17 26 17 26 17 9 16	21 26 13 6 10 19 26 11 17 26 27 12 26 27 12 26 27 12 34 22 14 17 20 17 13 16	4/VII (Cont to 8/VII	29 21 26 31 42 16 22 18 35 29 16 22 18 35 19 16 28 35 27 46 28	22 21 26 24 39 16 21 6 21 7 7 6 6 30 5 21 25 7 9 46		14/VI 18/VI 22/VI 26/VI	11 25 28 25 20 20 20 20 20 20 20 20 20 20 20 20 20	9 24 25 29 25 29 25 29 20 31 35 21 24 24 24 24 24 24 24 24 24 24 25 23 25 29 20 20 20 20 20 20 20 20 20 20 20 20 20	26/VI (Cont 30/VI	17 20 30 27 23 25 23 16 22 25 25 22 25 22 25 22 25 15 22 25 12	17 34 19 29 17 22 25 23 16 21 29 25 21 21 29 25 13 11 1	
4/vII	27 20 22 19 20	26 16 18 19 18 21 125 17 3 8 5 10 16 23 22 24 18 13	20/VII 24/VII 28/VII	46 29 15 47 20 45 20 15 20 20 15 20 10 20 20 10 20 20 10 20 10 20 10 20 10 20 10 20 10 20 10 20 10 20 10 20 10 20 10 20 10 20 10 20 10 20 10 20 10 20 10 20 10 20 20 20 20 20 20 20 20 20 20 20 20 20	$\begin{array}{c} 46\\ 23\\ 19\\ 6\\ 44\\ 16\\ 22\\ 3\\ 17\\ 12\\ 7\\ 9\\ 6\\ 11\\ 12\\ 13\\ 41\\ 12\\ 13\\ 12\\ 12\\ 12\\ 12\\ 12\\ 12\\ 12\\ 12\\ 12\\ 12$			16 34 40 15 30	14 33 35 12 29	8/VII 24/VII Total	6 24 21 20	1 6 22 16 18 13 1129	

Eggs hatched (or hatching) in 4 days Eggs not hatched in 4 days

Total 1936 1528

1958 Hatch Records

		-								-	
Date col- lected	No.egg scars	No. larvae	Date co l- lected	No. egg scars	No. larvae	Date co l- lected	No.egg scars	No . larvae	Date c ol- lected	No.egg scars	No. Larvae
11/VI 15/VI 19/VI 23/VI 27/VI 1/VII 5/VII	$\begin{array}{c} 7\\ 19\\ 15\\ 8\\ 17\\ 18\\ 28\\ 24\\ 32\\ 19\\ 33\\ 22\\ 32\\ 10\\ 21\\ 27\\ 19\\ 10\\ 31\\ 7\\ 27\\ 15\\ 44\\ 16\end{array}$	$\begin{array}{c} 7\\ 17\\ 13\\ 7\\ 16\\ 18\\ 24\\ 20\\ 30\\ 3\\ 28\\ 22\\ 22\\ 7\\ 16\\ 6\\ 17\\ 10\\ 26\\ 4\\ 18\\ 8\\ 43\\ 32\\ 8\end{array}$	9/VII (Contic 13/VII 17/VII 21/VII 25/VII 29/VII 2 VIII	$\begin{array}{c} 44\\ 9\\ 35\\ 15\\ 22\\ 7\\ 12\\ 21\\ 17\\ 13\\ 11\\ 14\\ 26\\ 10\\ 28\\ 38\\ 14\\ 30\\ 22\\ 9\\ 12\\ 6\\ 16\\ 6\end{array}$	42 8 24 17 7 19 42 15 36 6 6 6 6 6 2 28 28 4 5 4 6	11/VI 15/VI 19/VI 23/VI 27/VI 1/VII	24 25 25 20 5 6 20 5 9 6 33 12 46 25 25 27 10 10 13 27 25 27 10 10 12 27	$\begin{array}{c} 24\\ 22\\ 19\\ 4\\ 6\\ 25\\ 16\\ 12\\ 33\\ 17\\ 13\\ 26\\ 29\\ 25\\ 28\\ 9\\ 10\\ 9\\ 30\\ 19\\ 17\\ 17\\ 17\\ 17\\ 11\\ 13\\ 26\\ 29\\ 25\\ 28\\ 9\\ 10\\ 9\\ 30\\ 19\\ 17\\ 17\\ 17\\ 17\\ 17\\ 17\\ 17\\ 17\\ 17\\ 17$	5/VII (Cont 'd 9/VII 13/VII 13/VII 17/VII 25/VII 29/VII	$\begin{array}{c} 34\\ 34\\ 10\\ 10\\ 22\\ 10\\ 10\\ 22\\ 17\\ 10\\ 34\\ 10\\ 22\\ 34\\ 10\\ 36\\ 34\\ 36\\ 36\\ 31\\ 38\\ 58\\ 27\\ 17\end{array}$	32 35 12 59 17 13 40 14 20 39 58 30 24 35 7 47 24 7
			Total	997	775		42	36	Total	1280	1065

Eggs hatched (or hatching) in 4 days Eggs not hatched in 4 days

Ta	ble	6

Collection date	Associated with eggs	Associate with larg	ed No asso- vae ciation [*]	Apparently predatorized	Totals
1957			<u> </u>		
June 10 14 18 22 26 30 July 4 8 12 16	0 0 68 134 681 783 646 339 139 0	9 11 0 75 45 45 374 378 465 675	0 0 0 0 0 0 0 0 210	6 0 18 37 124 32 106 62 112	15 11 68 227 763 952 1052 823 666 997
Totals	0700	2077			
	2790	2077	210	497	5574
Percentages 1958	50:0	37.3	3.8	8.9	100.0
June 11 15 19 23 27 July 1 5 9 13 17	133 67 51 41 57 365 476 565 294 126	0 12 0 121 117 180 638 252 223 757	0 0 0 0 0 0 21 0 227	0 0 0 8 15 18 70 20 93	133 79 51 162 182 560 1132 908 537 1203
Totals	2175	2300	248	224	494 7
Percentages	44.0	46.5	5.0	4.5	100.0

Complete egg scar tally for portion of sampling period (Association becomes impractical after last dates given).

* The larvae from most of these egg scars had apparently completed feeding, or had been destroyed in later instars.

	<i>,</i>	Table	7
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	I In	star	пі	nstar	III	Instar	TV Instar		
Date	Scars			Larvae	Scars	Larvae	Scars	Larvae	
June 10	9	2	~~~			-			
22	28 47	24 28	-		-	-	-	-	
26	16	16	-	-	-	~	29	11	
30	בון בון	5 1	17	10	-	-	-	-	
July 4	51 43 64 50 53	16 21 30 39 48	21 45 23	3 29 9 -	2 22 - -				
8	21 37 39 15 45 21 25	13 6 3 5 6 15 4 10 12	27 17 15 25 -	20 13 1 11	48 	29		7	
12	34 41 19 -	1), 10 16 	31 50 18 20	7 36 15 16 -	18 28 44 -	6 11 33 -	33 31 36 25 20 17	12 10 17 5 13 5	
16	28		29 31 18 9 15 9 19 -	21 15 8 16 8 11 3 10 -	20 35 33 17 21 31 25 -	9 23 17 3 16 9 20	38 20 15 14 14 14 30 16 34 16 54	11 16 17 9 12 10 12 15 8 23	
20	24 1)4 1)4 -	18 3 14			14 16 22 20 -	12 1 19 7	16 44 28 54 16	5 28 23 40 15	

Numbers of egg scars and associated larvae for 1957

Table 7 (Cont'd)

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.

Date	I I Scars	I Instar Scars Larvae		Instar Larvae	III Scars	Instar Iarvae	TV I Scars	nstar Larvae
July 24		-	Scars -	-	5	5	10	6
28	6 14 24 30 26 9 30	2 13 17 28 21 4 11	12 16 20 8 17 17 5	11 8 10 4 12 16 5	21 16 27 23	21 14 10 19 -	19 19 	
Aug. 1	4 10 - -	2 14 	30 15 25 - -	16 11 10 - -	16 23 8 20 14 4	8 13 16 11 2	16 18 27 32	12 12 20 16
5	31	26	21	19	27	6	15	13
Totals	971	523	643	384	620	360	822	423

Table 8

	ΙI	nstar		instar	ΠI	Instar	IV I	instar
Date	Scars	Larvae	Scars	Larvae	Scars	Larvae	Scars	Larvae
June 15	12	11	-	-	-	-	-	-
23	28 9 36	15 7 20	13 22 13	6 19 9	-	-		-
27	6	6	26 14	19 10	40	26 -	בע 17	11 6
July l			20 27 33	14 13 20	29	2]4 		
5	16 19 22 19 108 45 23 26 35 42 25 29	5 16 1 19 68 18 7 13 20 1] ₄ 7 5	20 21 15 23	17 2 12 10 - -			59 20 17 19 11 -	

Numbers of egg scars and associated larvae for 1958

Table 8 (Cont'd)

	I Ir	star	II I	nstar	III	Instar	IV	Instar
Date	Scars	Larvae	Scars	Larvae	Scars	Larvae	Scars	Larvae
July 9	10 10 38 44 10 30 15	7 7 29 31 10 21 8	34		15 - - - - -	5	35	18
13	29 41 30 35 29	21 38 10 28 14	59	45 - - -	-	-		
17	13 22 30 25 15 28 15 12	11 20 19 14 9 27 14 10	29 21 13 52 - -	27 16 13 41 -		45 27 	98 33 25 -	36 28 13 18 - -
21	23 9 13 11 32 24 29 13 47 19 18	15 2 4 7 28 5 19 5 12 13	29 15 - - - -		23 35 24 26 21 - -	15 18 3 22 10 - -	24 70 10 5	19 40 10 2 -
25	19	יות ב	27 37 36	25 25 26	29 20 16	27 4 17	30 31 -	5 5 -
29	15 17 - -	5 15 - -	18 - -	16 - -	21 19 34 28 19	ר 8 14 18 7	32 20 15 -	16 9 14 -
Aug. 6	~	-	23	16	-	-	25	11
10	-	-	-	-	-	-	41	21
Totals	1281	786	640	<u>448</u>	515	289	684	340

	ΙI	Instar	TI	Instar	TII I	Instar	IV Ir	star	V Jr	nstar
Date	Reared	Collected	Reared	Collected	Reared	Collected	Reared	Collected	Reared	Collected
June 15 19 23 27	24 13 34 24	11 0 42 6	0 1 8 0 42	0 0 34 39	0 0 34	0 0 0 30	0 0 0 0	0 0 3	0 0 0 0	0 0 0 0
July 1 5 9 13 17 21 25 29	20 68 178 245 37 0 49 36	0 125 120 105 138 122 18 20	2 1 73 6 92 66 15 12	55 71 15 45 93 36 75 16	21 40 35 104 50 168 120 75	23 13 5 0 76 111 39 53	47 31 53 15 0 108 94 42	14 54 18 0 116 99 108 57	8 34 18 24 0 178 153 153	21 0 3 0 38 18 109 104
Aug. 2 6 10 14 18 22	1 10 0 0 0	0 0 0 0 0	19 15 0 0 0 0	0 0 16 0 0	6 0 15 0	6 0 0 0 0	52 5 0 1 0 0	1 15 23 3 0 0	110 24 39 25 16 1	78 53 47 24 4 0
Totals	 739	707	361	495	668	356	448	511	783	499

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Table 9

Numbers of larvae in field collections and in insectary rearings of previous collections

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TANTE TO	Ta	ble	10
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Date	9	Eggs	I Instar	II Insta r	III Instar	IV Instar	V Instar	Total Scars
June	10*	0 0	0 0	<1 <1	0 0	0 0	0 0	22 22
	14	12 12	0 0	0 0	0 0	0 0	0 0	12 12
	18	89 163	0 0	0 0	0 0	0 0	0 0	89 163
	22	752 1101	1 1	78 78	0 0	0 0	0 0	1324 2090
	26	4256 2914	26 26	0 0	0 0	12 12	0 0	5945 3672
	30	10393 6827	2 3	10 10	0 0	0 0	0 0	11591 6585
July	4	6804 7354	710 42	74 93	12 12	0 0	0 0	16023 13261
	8	885 992	69 12	40 50	75 90	4 4	<1 <1	1254 830
	12	795 392	50 66	254 83	357 126	240 35	2 2	23143 1354
	16	0 0	43 44	147 272	218 216	330 2415	79 37	10755 4355
	20	872 198	36 53	1 2	28 50	208 511	789 1036	16687 22603
	24	229 29	0 0	0 0	2 3	3 3	50 88	4431 589
	28	212 300	407 416	313 39	82 72	13 22	64 24	9026 10950
Aug.	1	0 0	2 2	170 177	452 644	537 448	648 28	30279 17338
	5	0 0	68 68	26 26	256 314	385 394	930 1012	42866 10910
	9	0 0	0 0	0 0	0 0	< 1 < 1	23 71	19640 4072

Inter- and intra-tree mean squares (1957 data)

*The first number in each pair is the inter-tree mean square, the other is the intra-tree mean square.

Table 1	11
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Date [*]	Eggs	I Instar	II Instar	III Instar	IV Instar	V Instar	Total Scars
June 11	237	0	0	0	0	0	237
	-14	0	0	0	0	0	く1
	117	12	0	0	0	0	175
15	0	0	0	0	0	0	0
	0	0	0	0	0	0	0
	29	0	0	0	0	0	29
19	116	0	0	0	0	0	116
	0	0	0	0	0	0	0
	87	0	0	0	0	0	87
23	0	86	116	0	0	0	885
	0	0	0	0	0	0	0
	325	4	86	37	0	0	1475
27	0	0	< 1	<1	1	0	29
	0	0	1	4	4	0	149
	706	0	24	53	20	49	1946
July 1	2716	0	51	0	0	0	2930
	1396	0	10	0	0	0	4704
	3884	698	46	14	228	0	22301
5	372 0 0	0 0 5	. 0 4	0 0 0	0 0 0	0 0 1	368 -23 208
9	3435	344	22	0	32	0	8431
	680	269	0	0	0	0	3277
	293	443	6	0	0	0	1992
13	2771	20	202	0	0	0	2651
	506	42	0	0	0	0	2557
	309	167	217	200	116	116	17473
17	143	173	171	73	186	2	3120
	0	23	16	-9	-12	0	3050
	90	171	36	143	236	22	7649

Estimated variances and covariances for 1958

Table 11 (Contid)

Date	Eggs	I Instar	II Instar	III Instar	IV Instar	V Instar	Total Scars
July 21	44	47	17	8	2	<1	2178
	0	0	0	24	_<1	0	5685
	51	0	62	72	3	43	21256
25	73	32	101	4	389	253	18077
	0	26	0	2	0	18	5046
	29	23	0	35	32	58	4701
29	بلا	0	24	38	77	283	4269
	0	0	0	0	0	_l	-440
	0	0	0	0	0	2	2020
Aug. 2	78	0	0	4	<1	135	11005
	50	0	0	0	○	5	752
	32	0	0	0	<1	5	1128
6	0	0	0	0	12	79	3865
	0	0	0	0	24	4	5613
	0	0	26	0	48	5	12826
10	0	0	0	0	0	102	7953
	0	0	0	0	0	6	7455
	0	0	0	0	<1	11	11530
זע	0	0	0	0	0	<1	1634
	0	0	0	0	0	0	-519
	0	0	0	0	0	0	.1979
18	0 0 0	0 0 0	0 0 0	0 0 0	0 0 0		1212 1246 9657

* The first figure in each group of three is the variance for the random half of the previous sample, the third is the variance for the overlapping half of the sample for the date given, and the middle figure is the covariance between the two.